

Molecular Method Development to Identify Foodborne *Sarcocystishominis* in Raw Beef Commercial Hamburger

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Background: *Sarcocystis* spp. is zoonotic parasitic pathogen endangering safety of meat and derived meat products such as hamburgers which is among the most popular fast foods worldwide.

Objectives: The current study aimed to design a protocol for molecular identification of *Sarcocystis hominis* in commercial hamburgers using PCR-RFLP with target of 18S rRNA.

Materials and Methods: A total of 25 raw commercial hamburger samples were randomly collected from supermarkets of Yazd city, Iran. Five mm slices from different parts of each sample were selected, well mixed, and then preserved in ethanol 70% at -20°C for the next steps. The genomic DNA was extracted using salting out method. Detection and identification of *Sarcocystis* isolates were performed using PCR-RFLP. The 18S rRNA gene sequence was mined from GenBank and the specific primer pair was designed using Primer3 software. Restriction fragment length polymorphisms (RFLP) analysis was performed using *Bfal* and *Rsal* restriction enzymes. The digestion was analyzed, using agarose gel electrophoresis alongside 100base pair DNA ladder.

Results: Among 25 commercial hamburger samples, 17 samples showed a PCR product around 900 bp which could detect *Sarcocystis* spp. After RFLP with *Bfal*, the restriction fragments of 376 bp and 397 bp detected *S. hominis* or *S. hirsuta* and fragments of 184 bp, 371 bp and 382 bp detected *S. cruzi*. After RFLP with *Rsal*, the restriction fragments of 376 bp and 557 bp detected *S. hirsuta* and fragment of 926 bp, without any digestion, detected *S. hominis*. For verification, each species detected in samples was randomly selected and sent for sequencing and the results were analyzed with BLAST.

Conclusions: In conclusion, the current study developed a practical technique to detect the prevalence of *S. hominis* in meat products such as hamburgers.

Keywords: *Sarcocystis* spp.; Food borne Diseases; Molecular Diagnostic Techniques

1. Background

Sarcocystis spp. as the obligatory intracellular two-host parasites belong to the family of Sarcocystidae and phylum of Apicomplexa (1, 2). Among the numerous species, just *S. hominis* and *S. suihominis* can infect humans, which cattle and pigs are their intermediate hosts, respectively. Intestinal infection in human may occur by consumption of undercooked beef or pork containing the larvae stage named sarcocyst (3-5). The clinical signs are digestive system disorders including nausea, vomiting, stomachache and diarrhea (3, 4). *Sarcocystis* spp. is considered as one of the important opportunistic agent in acquired immune deficiency syndrome (AIDS) patients (6). *Sarcocystis* spp. is zoonotic parasitic pathogens endangering safety of meat and derived meat products such as hamburgers which are among the most popular fast foods worldwide. Each year, about five billion hamburgers are consumed just in

the USA (7). Although, there is no exact information regarding per capita of meat products in Iran, it is estimated that the annual consumption of hamburgers is considerable in this country. As beef is the main ingredient of hamburgers in Iran, the occurrence of *Sarcocystis* spp. in this product should be considered. The routine detection of these parasites includes pathological and serological methods. Gross examination, impression smears, and digestion methods are the usual methods in his to pathological methods and indirect fluorescent antibody test (IFAT), and Enzyme-linked immunosorbent assay (ELISA) are used in serological methods (8). As, these methods are either time-consuming or non-sensitive enough to identify *Sarcocystis* species in some methods with similar morphology, therefore, the techniques based on DNA analysis will be useful to identify any organisms (9), also

Sarcocystis species. The variable regions of the 18S rRNA gene provide useful targets to identify and characterize different species, even from the same genus (10). It is found that such sequences for *Sarcocystis* show low inter-specific homology relative to intraspecific comparisons.

2. Objectives

As *S. hominis* is the most important species transmitted by cattle in Iran, the current study aimed to design a protocol for molecular identification of *Sarcocystis hominis* in commercial hamburgers using PCR-RFLP targeting 18S rRNA.

3. Materials and Methods

3.1. Sampling

A total of 25 raw frozen commercial hamburger samples with different meat content, from some factories were randomly purchased from supermarkets of Yazd city, Iran. In the laboratory, all samples were cut into 5 mm slices and examined carefully by naked eye for the presence of macro cysts of *Sarcocystis*. Five slices from different parts of each hamburger sample were selected and well mixed, and then preserved in ethanol 70% at -20°C for the next steps.

3.2. DNA Extraction

The genomic DNA was extracted using salting out method (11). Approximately 30mg of each sample was suspended in 900µL NET (NaCl, 50 mM; EDTA pH 8, 25 mM, Tris-HCl pH 7.8, 50 mM) buffer, with 10 µL proteinase K (20 mg/mL, Fermentas, EO049), and Sodium dodecyl sulfate (SDS) with the end concentration of 1% in order to lysis the sample, followed by rapid mixing and incubation at 56 °C for an overnight. The purification of DNA was performed by adding 250 µL 6M NaCl. After centrifugation, the supernatant was transferred to a new sterile 1.5 mL micro tube. The DNA precipitation was performed by adding a double volume of chilled absolute ethanol. After washing with ethanol 70%, the DNA pellet was dried and re-suspended in 100µL of sterile double distilled water and stored at -20°C till next steps.

3.3. Detection and Identification

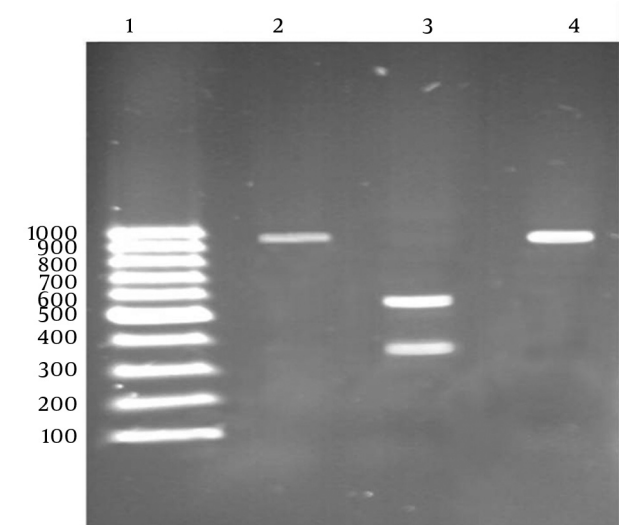
Detection and identification of *Sarcocystis* isolates were performed using PCR-RFLP based on the authors' previous study (11). The 18S rRNA gene sequence was mined from genbank and the specific primer pair was designed using Primer3 software that included sarF 5'-CGT GGT AAT TCT ATG GCT AAT ACA -3' and sarR 5'-TTT ATG GTT AAG ACT ACG ACG GTA-3'. The amplicon size was estimated around 900 base pair. Amplification was performed using 1X PCR buffer, 1.5 mM MgCl₂, 0.2 mM each dNTP, 1U of Taq DNA polymerase, 10 pmol of each primer and 100ng of the genomic DNA as a template. This amplification was

performed for the negative control including all the reagents but not the template. The amplification program was done with an initial denaturation of 94°C for five minutes, followed by 30 cycles of 94°C for 60 seconds, 58 °C for 60 seconds and 72°C for 60 seconds and it was finalized with the extension of 72°C for five minutes. The amplicon was analyzed on 1% agarose gel using electrophoresis, alongside with 100 bp of the DNA ladder. RFLP analysis was performed using *Bfal* and *RsaI* restriction enzymes (11). The reaction was carried out with 10U either *Bfal* or *RsaI* restriction enzyme, 1X specific buffer and 10µL of the PCR products, then, was incubated for 16 hours at 37°C according to the manufacturer's recommendations. The digestion was analyzed, using agarose gel electrophoresis alongside the 100 bp DNA ladder.

4. Results

Among 25 commercial hamburger samples, 17 (%68) showed a PCR product around 900 bp which could detect *Sarcocyst Spp*. RFLP analysis was performed using *Bfal* and *RsaI* restriction enzymes and it was analyzed using the agarose gel electrophoresis. After, RFLP with *Bfal*, the restriction fragments of 376 bp and 397 bp detected *S. hominis* or *S. hirsuta* and fragments of 184 bp, 371 bp and 382 bp detected *S. cruzi*. The restriction enzyme of *RsaI* was used in order to distinguish between *S. hominis* and *S. hirsuta*. After RFLP with *RsaI*, the restriction fragments of 376 bp and 557 bp detected *S. hirsuta* and fragment of 926 bp, without any digestion, detected *S. hominis* (Figure 1). For verification, each species detected in the samples were randomly selected and sent for sequencing and the results were analyzed with BLAST.

Figure 1. PCR-RFLP to Identify *S. hominis*



Lane 1, 100 bp DNA ladder; Lane2, digestion with *RsaI*, no digestion; Lane3, digestion with *Bfal* resulting two fragments of 376bp and 550bp; Lane4, PCR product before any digestion with 926 bp in size.

5. Discussion

The family *sarcocyst* includes cyst-forming coccidia, which parasitize a wide variety of vertebrates including humans. *Sarcocystis* species are among the most common and widespread protozoan parasites of livestock with economic importance around the world. A large number of vertebrate species serve as intermediate or definitive hosts for *Sarcocystis* species, and some hosts are often infected by several different *Sarcocystis* species at the same time. Such mixed infections lead to taxonomic confusion within the genus *Sarcocystis*. This confusion is compounded by incomplete knowledge of the life cycles of many *Sarcocystis* species and the paucity of informative morphological characters for apicomplexans in general. The species are distinguished by the structure of the cyst wall under light and transmission electron microscopies (12).

In all previous surveys, impression smear, histological or digestion techniques were used to detect *Sarcocystis* spp. Since none of the indicated assays could identify *S. hominis* and *S. hirsuta* (13, 14), the prevalence of these species in the cattle population of Iran was not reported earlier. Therefore, designing and development of a sensitive and specific protocol could have an important role to detect and identify *Sarcocystis* spp. and report the true prevalence of this zoonotic pathogen (11). The current study used 18S rRNA as a suitable target because this region has little interspecific variation but high sequence divergence between the species (15). This variable in the region could make it suitable to identify species. Previously, there were some documents for molecular detection and identification (16, 17) but the current design introduced just two restriction enzymes for identification that is cost-effective especially for developing countries (18, 19). As shown in the results, the designed primer could amplify an amplicon around 900 bp and the restriction digestion analysis with *BfaI* showed 376 bp and 397 bp restriction fragments for *S. hominis* or *S. hirsuta* and 184bp, 371 bp and 382 bp for *S. cruzi*. The restriction enzyme of *RsaI* was used in order to distinguish between *S. hominis* and *S. hirsuta*. After RFLP with *RsaI*, 376 bp and 557 bp restriction fragments detected *S. hirsuta* and 926 bp fragment, without any digestion, detected *S. hominis*. In conclusion, the current study developed a practical technique which could detect prevalence of *S. hominis* in meat products such as hamburgers.

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References

1. Nematollahia A, Khoshkarder A, Helan JA, Shahbazi P, Hassanzadeh P. A study on rate of infestation to *Sarcocystis* cysts in supplied raw hamburgers. *J Parasit Dis*. 2013;1-4.
2. Oryan A, Sharifiyazdi H, Khordadmehr M, Larki S. Characterization of *Sarcocystis fusiformis* based on sequencing and PCR-RFLP in water buffalo (*Bubalus bubalis*) in Iran. *Parasitol Res*. 2011;109(6):1563-70.
3. Fayer R. *Sarcocystis* spp. in human infections. *Clin Microbiol Rev*. 2004;17(4):894-902.
4. Rahdar M, Salehi M. The prevalence of *Sarcocystis* infection in meat-production by using digestion method in Ahvaz. *Iran Jundi J Microbiol*. 2011;4:36-45.
5. Bucca M, Brianti E, Giuffrida A, Ziino G, Cicciari S, Panebianco A. Prevalence and distribution of *Sarcocystis* spp. cysts in several muscles of cattle slaughtered in Sicily, Southern Italy. *Food Control*. 2011;22(1):105-8.
6. Velasquez JN, Di Risio C, Etchart CB, Chertcoff AV, Mendez N, Cabrera MG, et al. Systemic sarcocystosis in a patient with acquired immune deficiency syndrome. *Hum Pathol*. 2008;39(8):1263-7.
7. Prayson B, McMahon JT, Prayson RA. Fast food hamburgers: what are we really eating? *Ann Diagn Pathol*. 2008;12(6):406-9.
8. Dubey JP, Speer CA, Fayer R. *Sarcocystosis of animals and man*. CRC Press Inc; 1989.
9. Amini-Bavil-Olyae S, Pourkarim M. How can a novel molecular diagnostic assay instill confidence in researchers and encourage its future use? *Hepat Mon*. 2012;12(4):292-3.
10. Neefs JM, Van de Peer Y, De Rijk P, Goris A, De Wachter R. Compilation of small ribosomal subunit RNA sequences. *Nucleic Acids Res*. 1991;19 Suppl:1987-2015.
11. Zohourtabar A, Mehrizi SR. First molecular identification of *Sarcocystis hirsuta* in Iranian beef: A case report. *J Food Qual Hazards Control*. 2014;1:32-4.
12. Jeffries AC, Schnitzler B, Heydorn AO, Johnson AM, Tenter AM. Identification of synapomorphic characters in the genus *Sarcocystis* based on 18S rDNA sequence comparison. *J Eukaryot Microbiol*. 1997;44(5):388-92.
13. Jehle S, van Rossum B, Stout JR, Noguchi SM, Falber K, Rehbein K, et al. alphaB-crystallin: a hybrid solid-state/solution-state NMR investigation reveals structural aspects of the heterogeneous oligomer. *J Mol Biol*. 2009;385(5):1481-97.
14. Esteller More E, Pons Calabuig N, Romero Vilarino E, Puigdollers Perez A, Segarra Isern F, Matino Soler E, et al. [Dentofacial development abnormalities in paediatric sleep-related breathing disorders]. *Acta Otorrinolaringol Esp*. 2011;62(2):132-9.
15. Boughattas S, Salehi R. Molecular approaches for detection and identification of foodborne pathogens. *J Food Qual Hazards Control*. 2014;1:1-6.
16. Shekarforoush SS, Razavi SM, Abbasvali M. First detection of *Sarcocystis hirsuta* from cattle in Iran. *Iran J Vet Res Shiraz Univ*. 2013;14(2):155-7.
17. Hamidinejat H, Razi Jalali MH, Nabavi L. survey on sarcocystis infection in slaughtered cattle in south-west of iran, emphasized on evaluation of muscle squash in comparison with digestion method. *J Anim Vet Adv*. 2010;9(12):1724-6.
18. Hajimohammadi B, Dehghani A, Ahmadi Moghadam M, Eslami G, Oryan A, Khamesipour A. Prevalence and species identification of *Sarcocystis* in raw hamburgers distributed in Yazd, Iran using PCR-RFLP. *J Food Qual Hazards Control*. 2014;1:15-20.
19. Hajimohammadi B, Dehghani A, Moghaddam Ahmadi M, Eslami G, Oryan A, Yasini Ardakani SA, et al. Isolation of *S. hirsuta* from traditional hamburger of Iran by PCR-RFLP. *J Isfahan Med School*. 2014;32(273).